

Cleaning Oak Ridge 15mL Reusable Centrifuge Tubes (Improved Fuhrman lab protocol)

Last updated: 2025-03-03 by J. McNichol

1. Using water and diluted Alconox/Sparkleen or other lab solid soap (1-2%), thoroughly scrub tubes with the scrubber brush to remove any residual DNA. Be sure to scrub the side/bottom of the tube where the DNA precipitates during centrifugation.
2. Rinse each tube AT LEAST 3x with regular distilled water, or more if necessary, to remove all soap. No bubbles should form once the tube is clean. Make sure to rinse the lid as well.
3. Put tubes into an autoclavable rack. Autoclave for 30 minutes on the dry cycle. Make sure lids are loose, otherwise the tubes will get warped/melt.
4. After cooling, fill each tube with 5 mL 5% HCl, close lids completely, shake gently, and allow to sit for at least 12 hours at room temperature.
5. The next day, dump out the acid and rinse each tube 3x with 5mL **molecular grade water** (only use the bottle labelled for rinsing).
6. Flush rinse with 5-10 mL of 100% ethanol to aid in removal of water and drying (enough to cover all inner surfaces of tube).
7. Let air-dry upside down in the CleanSpot in styrofoam racks (ask if you're not sure how this works). Do not place in drying rack. Lids should be dried face-up on the shelf on a clean surface (tin foil or kimwipe).
8. Once fully dry (usually the next day), UV both caps and tubes on top shelf of CleanSpot for 20 minutes, then cap and set aside until use in DNA extraction.